Louisiana Office of Public Health Laboratories	
Test Name	Arbovirus Panel
PHL Location	Office of Public Health Laboratory Baton Rouge
CPT Code	86788, 86653
Synonyms	WNV, SLE, EEE, Flavivirus, Mosquito borne encephalitis, Viral encephalitis
	Prior notification to Infectious Disease Epidemiology required: Email: <a href="mailto:christine.scott-waldron@la.gov">christine.scott-waldron@la.gov</a> Phone: 504-568-8301 24 hour cell: 800-256-2748
Brief Description of Test	Arbovirus panel includes.  Samples received, collected within 7 days of symptom onset get PCR for WNV and WNV/SLE and EEE IgM  Samples received, collected > 7 days of symptom onset get
	WNV/SLE and EEE IgM  WNV PCR currently being performed for Epi use only, no patient report will be issued.
Possible Results	See each test for possible results
Reference Range	Negative
Specimen Type	Serum or CSF
Specimen Container(s):	Red top tubes, Marble top tubes, polypropylene vials
Minimum volume accepted:	1 mL of serum 1.5 mL of CSF
Collection Instructions	Specimens (serum or CSF) should only be collected by personnel that have been properly trained. Care should be taken during specimen collection and handling to avoid generation of aerosols. Serum should be collected in a plastic tube, such as a vacutainer, which does not contain an anticoagulant. The collection tube may or may not contain a serum separator. If collected in a tube without serum separator, serum must be aliquoted into screw cap tubes before shipment to laboratory. Depending on the type of collection tube, the amount of time it will take for the blood to clot could take up to 60 minutes. Separation of serum from cells should take place within 2 hours of collection to prevent erroneous test results according to NCCLS guidelines.

	Follow the package insert for the collection tube you use.
	Label specimen with Patient Name and a 2 <sup>nd</sup> unique identifier such as a chart number or medical record number. DOB is not considered unique.
	Complete a Lab Form 96 to accompany the serum or CSF sample. Lab submission form must be thoroughly completed with patient's first and last name, 2 <sup>nd</sup> patient identifier, gender, date of birth, date of collection, time of collection, test requested, and submitter's name, address, and contact number.
	For this assay, date of symptom onset is requested.
	Two unique identifiers <b>MUST</b> be recorded on the tube <b>AND</b> the Lab 96 form. A missing identifier on the tube will be an automatic rejection. If the identifiers are missing from the Lab 96 form, the submitter must be contacted and a new form with this information must be faxed back to the lab before testing will take place.
Storage and Transport Instructions	Specimens should be shipped refrigerated (2-8°C). Specimens can be stored for up to 1 month at 2-8°C. For longer storage, specimens must be frozen at -20°C or colder after collection. Frozen specimens must be shipped on dry ice. If samples are frozen, document the date and time the sample was frozen.
Causes for Rejection	Hemolyzed, lipemic, or icteric specimens must be rejected. Improper labeling, expired collection tubes, unspun samples, tubes that contain less than 90% of the total drawing capacity (QNS), specimen age >1 month if specimen has not been frozen at -20°C or colder. Improper storage and improper transport temperature requirements are also reasons for rejection.
Limitations of the Procedure	See each test for procedure limitations
Interfering Substances	Grossly hemolyzed, lipemic, or icteric specimens
	Duplex Microsphere-Based Immunoassay for Detection of Anti-West Nile Virus and Anti-St. Louis Encephalitis Virus Immunoglobulin M Antibodies", <u>Clinical and Diagnostic Laboratory Immunology</u> , May 2005, p. 566-574
References	Validation of a Microsphere-Based Immunoassay for Detection of Anti-West Nile Virus and Anti-St. Louis Encephalitis Virus Immunoglobulin M Antibodies", <u>Clinical and Diagnostic Laboratory Immunology</u> , September 2007, p. 1084-1093
	Protocols from CDC for EEE IgM MIA testing
	Protocols from CDC for West Nile Virus PCR

Additional Information	See each test for additional information.
Release Date	03/15/2016

Warning: If you have printed a copy of this information please be advised that the Louisiana Office of Public Health Laboratories website and methods are updated on a regular basis. Please check the on-line version of this document to ensure you are relying on the most recent release.

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